

Neuronal Cell Death and Synaptic Pruning driven by Spike-Timing Dependent Plasticity

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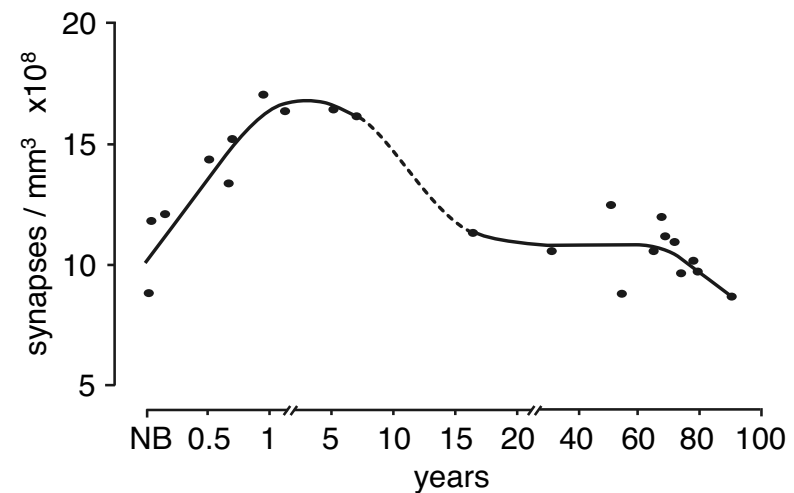
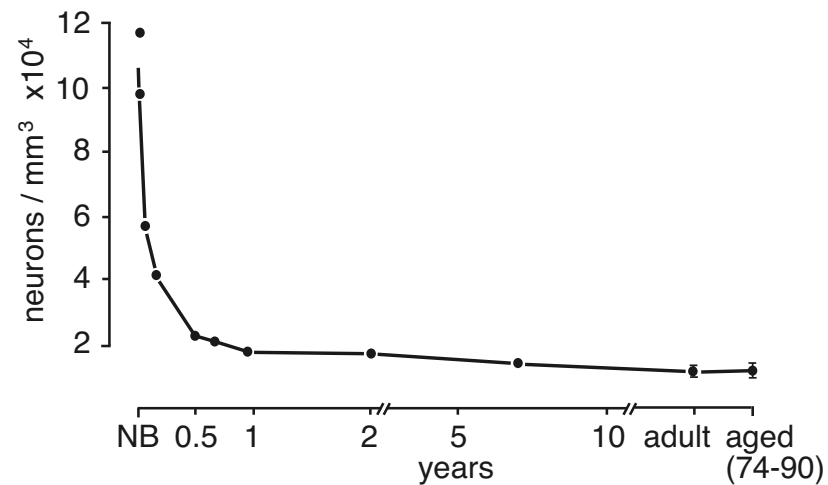
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introduction: synaptogenesis and synaptic pruning

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modified from Huttenlocher, *Synaptic density in human frontal cortex – developmental changes and effects of aging*, Brain Research, 163:195–205, 1979

- The memory performance of a network is optimally maximized if, under limited metabolic energy resources restricting their number and strength, synapses are first overgrown and then pruned.

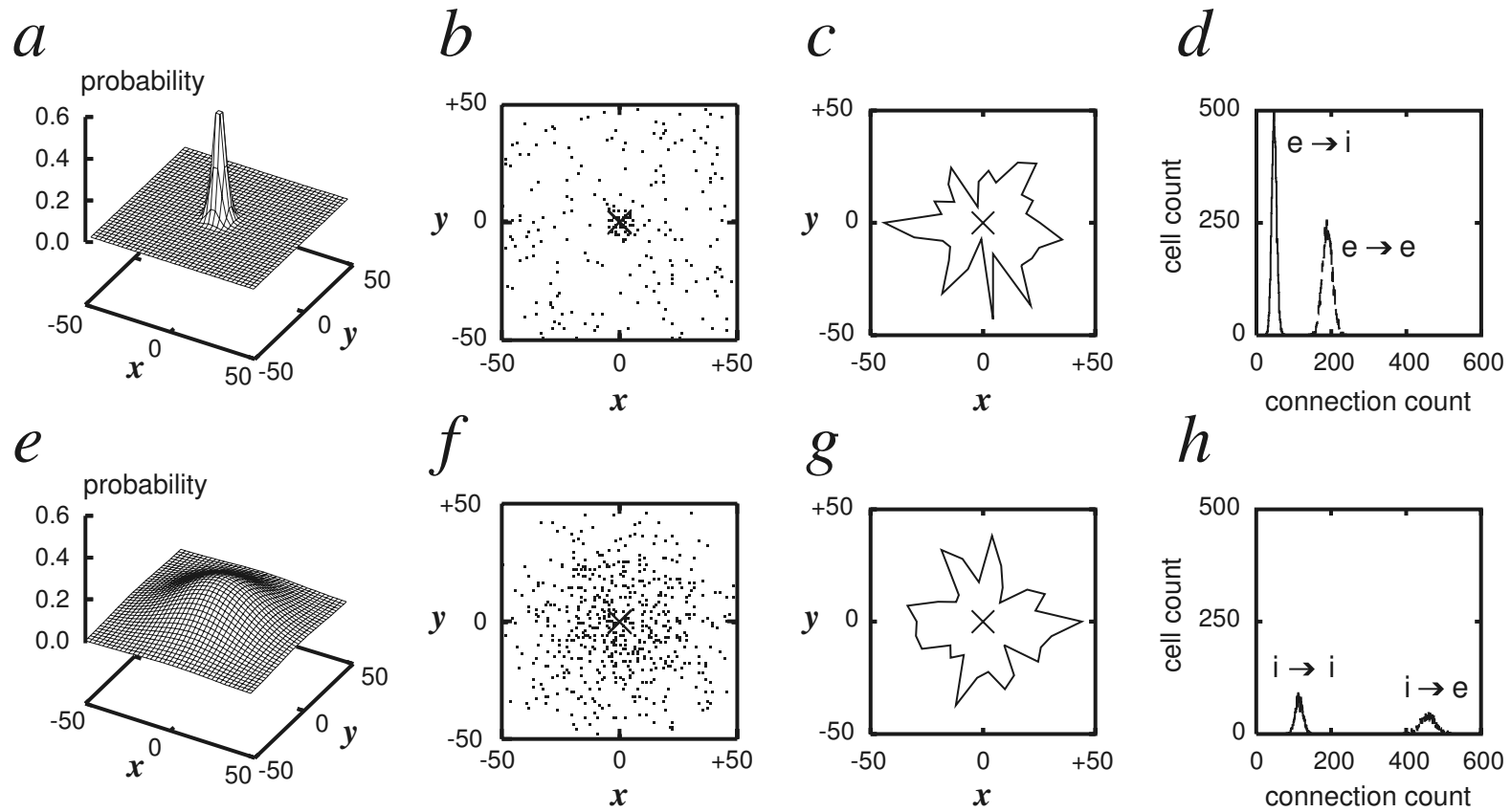
Chechik *et al.*, *Synaptic pruning in development: A computational account*, Neural Computation, 10(7):1759–77, 1998

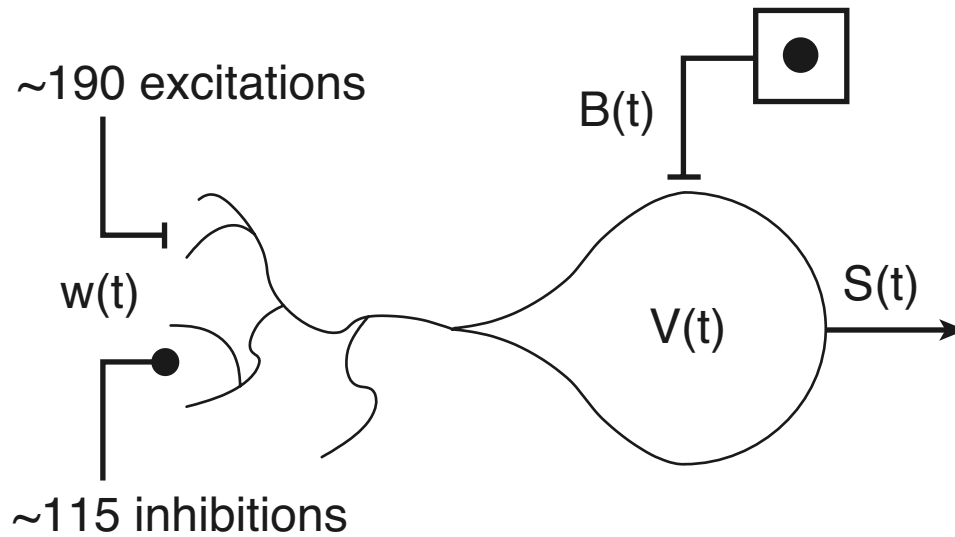
- Neuronal regulation might maintain the memory performance of networks undergoing synaptic degradation.

Horn *et al.*, *Memory maintenance via neuronal regulation*
Neural Computation, 10(1):1–18, 1998

- STDP has been shown to maintain the postsynaptic input field.

Abbott *et al.*, *Synaptic plasticity: taming the beast*
Nature Neuroscience, 3:1178–83, 2000





<i>Type I</i>	=	excitatory 80%
<i>Type II</i>	=	inhibitory 20%
V_{rest}	=	-76 [mV]
θ_i	=	-40 [mV]
τ_{mem}	=	15 [ms]
t_{refract}	=	exc:3 inh:2 [ms]
λ_i	=	5 [spikes/s]
n	=	50

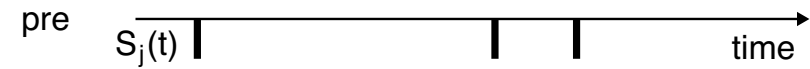
$$V_i(t+1) = V_{\text{rest}[q]} + (1 - S_i(t)) \cdot ((V_i(t) - V_{\text{rest}[q]}) \cdot k_{\text{mem}[q]}) + \sum_j w_{ji}(t) + B_i(t)$$

$$S_i(t) = \mathcal{H}(V_i(t) - \theta_{q_i})$$

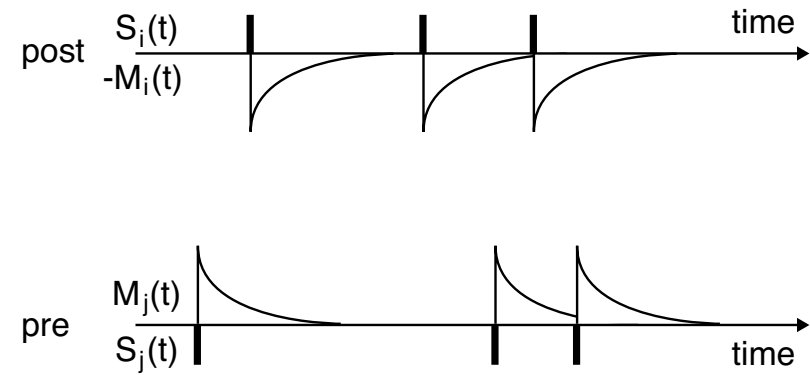
$$w_{ji}(t+1) = S_j(t) \cdot A_{ji}(t) \cdot P_{[q_j, q_i]}$$

$$B_i(t+1) = \mathcal{P}_{\text{reject}}(\lambda_{q_i}) \cdot n \cdot P_{[q_1, q_i]}$$

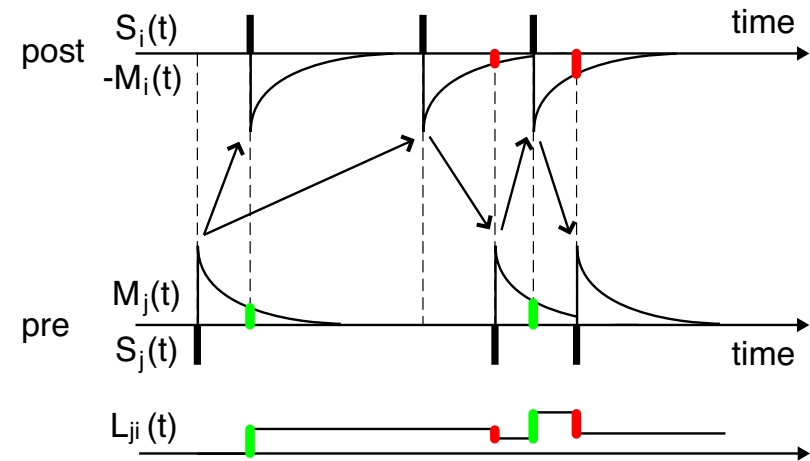
$$L_{ji}(t + 1) = L_{ji}(t) \cdot k_{\text{act}[q_j, q_i]} \\ + (S_i(t) \cdot M_j(t)) \\ - (S_j(t) \cdot M_i(t))$$



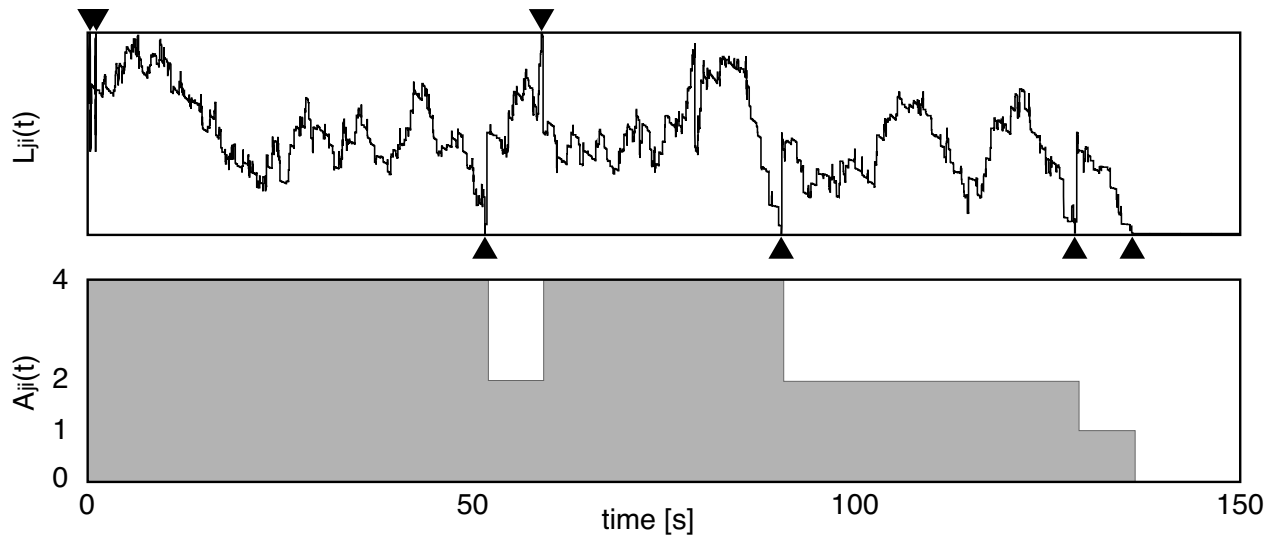
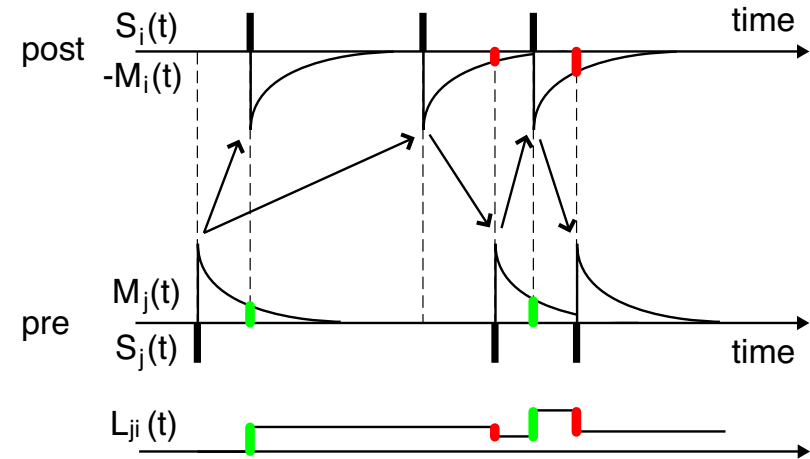
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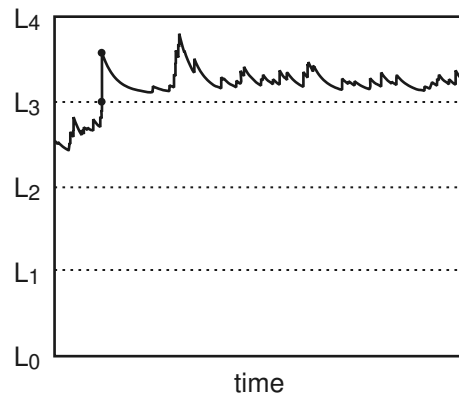
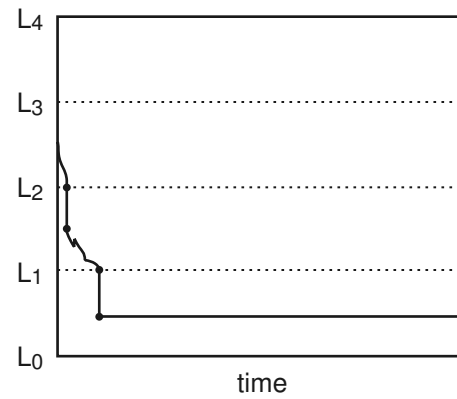
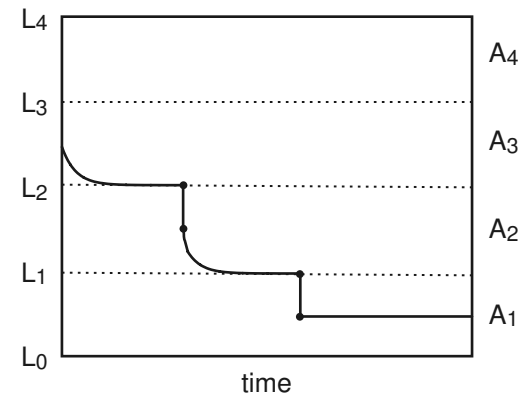
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$$L_{ji}(t+1) = L_{ji}(t) \cdot k_{\text{act}[q_j, q_i]} + (S_i(t) \cdot M_j(t)) - (S_j(t) \cdot M_i(t))$$



$$w_{ji}(t+1) = S_j(t) \cdot A_{ji}(t) \cdot P_{[q_j, q_i]}$$

a*b**c*

Strongly Interconnected (SI) units

at the end of the simulation

set of cells (discarding input units)

maintaining $k_{out} \geq 3$ and $k_{in} \geq 3$

with strongest activation level ($A_{ji}(t) = 4$)

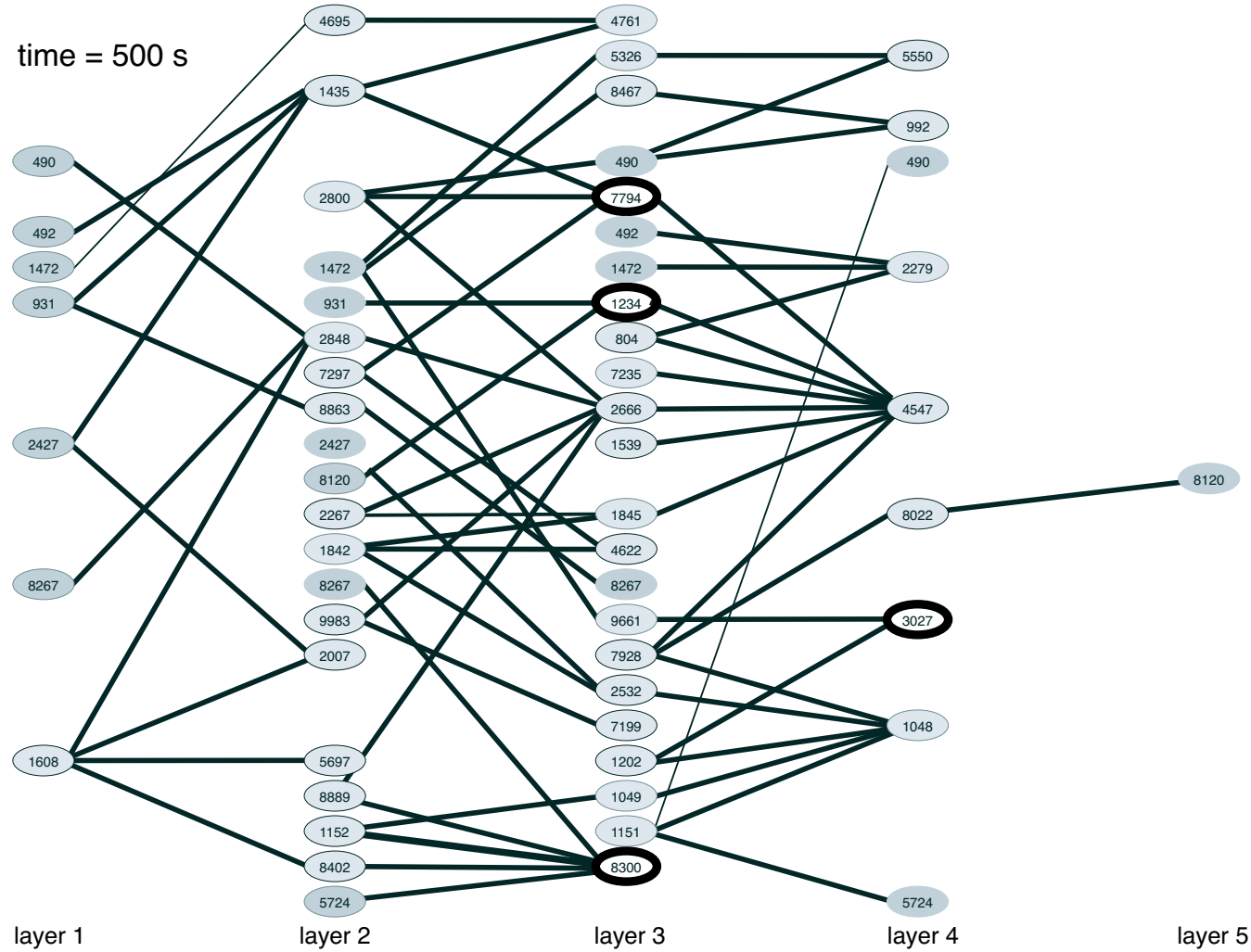
with units with the same properties.

Neighbourhood

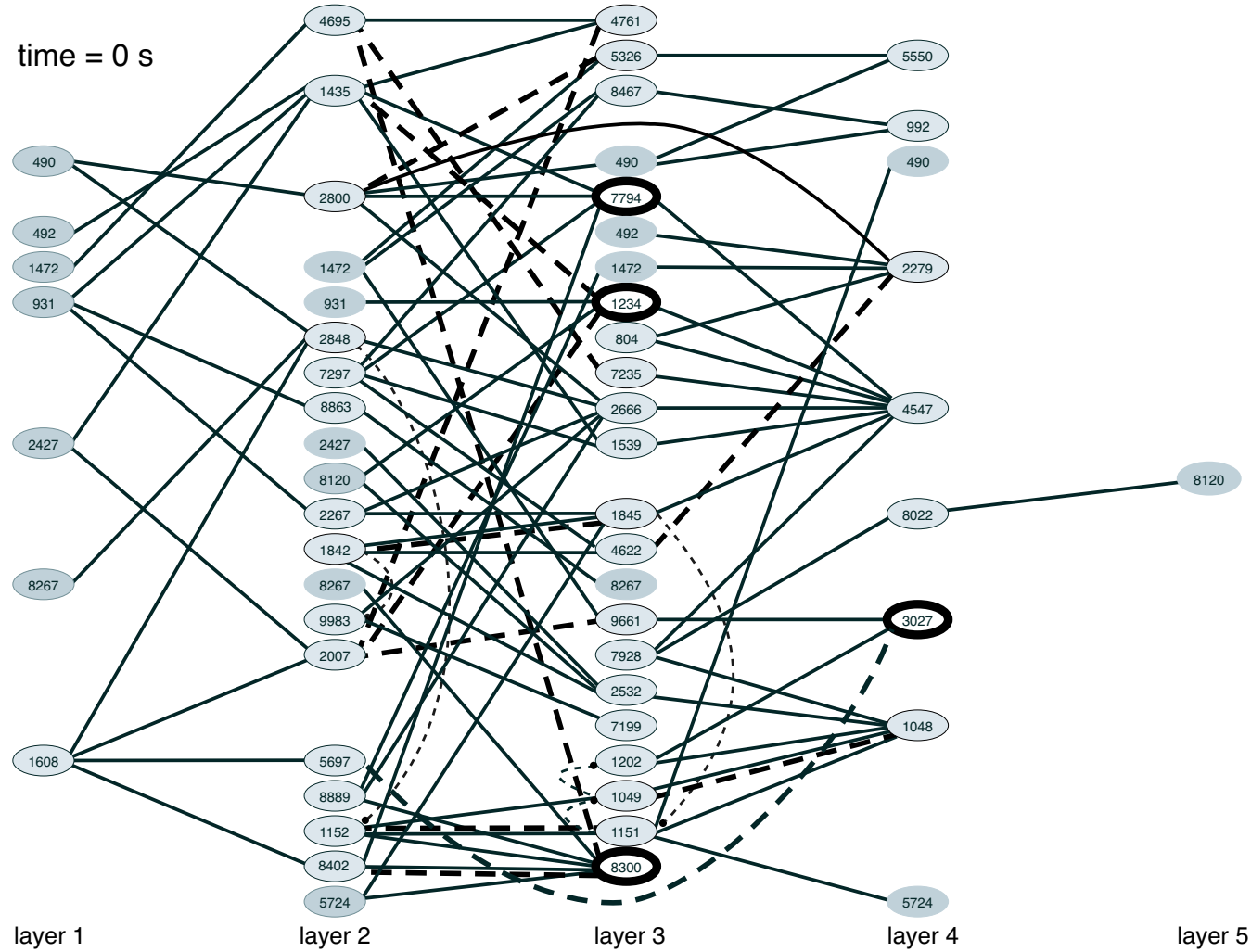
all excitatory units (including input units)

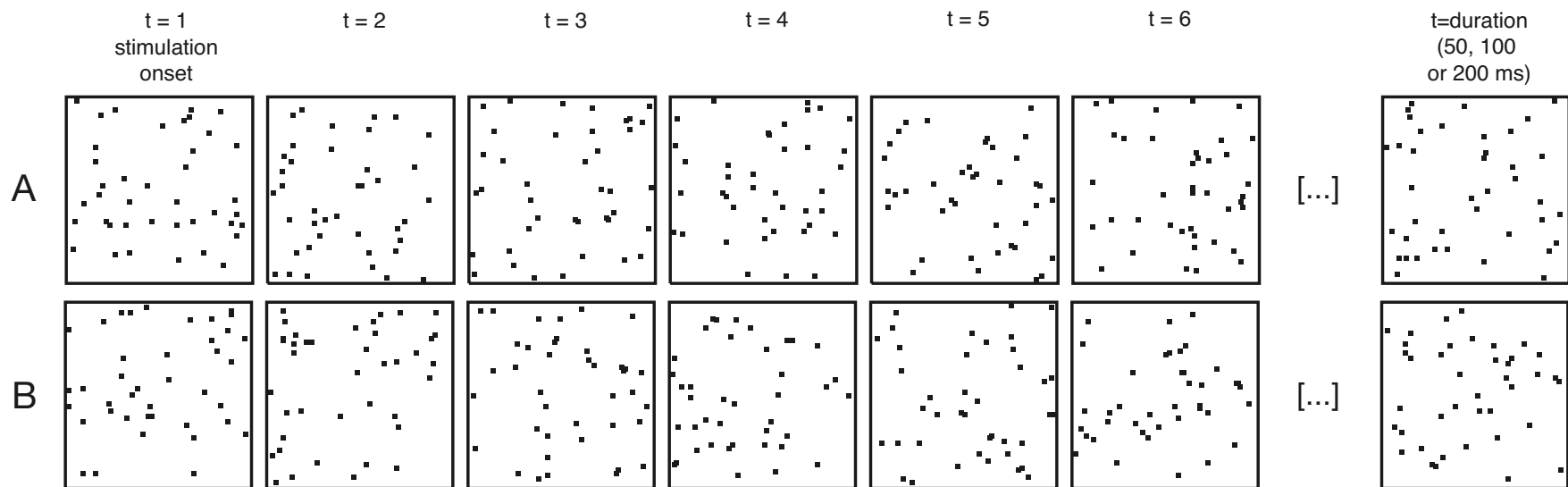
with at least one projection to or from SI-units.

Iglesias et al., *Emergence of Oriented Cell Assemblies*
 Associated with Spike-Timing-Dependent Plasticity, LNCS 3696:127-132, 2005



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 Associated with Spike-Timing-Dependent Plasticity, LNCS 3696:127-132, 2005



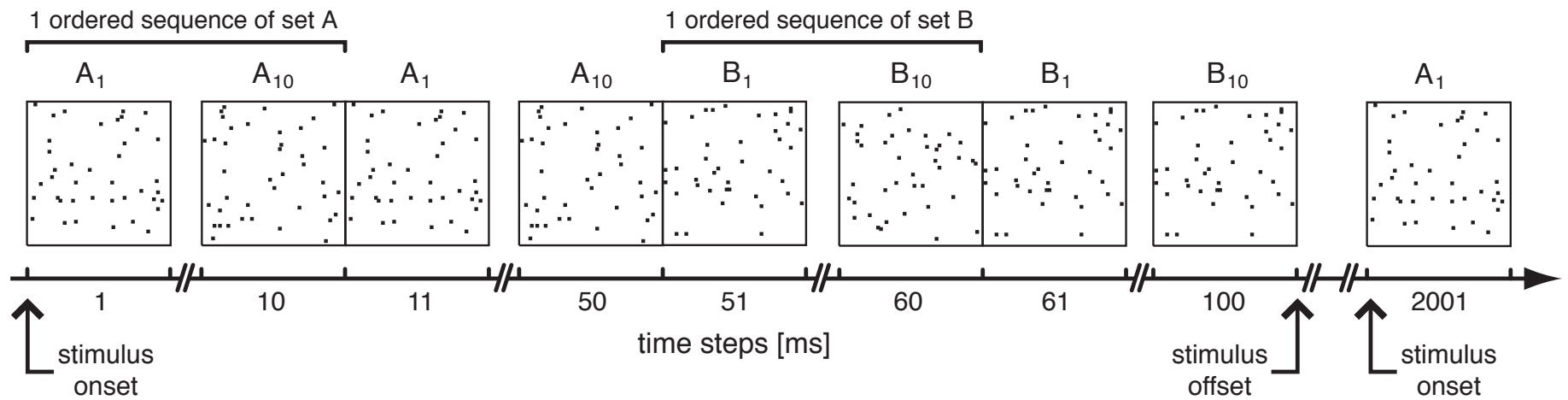


every 2 seconds

10 groups of 40 units activated in sequence (10% input units)

during 100 time steps ($5 \times A + 5 \times B$ or $5 \times B + 5 \times A$)

Animated sequences are available for both stimuli [A](#) and [B](#).



- **apoptosis induced by excessive firing rate**

50 ms running window

firing rate threshold:

$$\theta_{\nu}^{exc} = 245 \text{ sp/s for excitatory units}$$

$$\theta_{\nu}^{inh} = 250 \text{ sp/s for inhibitory units}$$

death probability function: $\mathcal{P}_{\text{apopt}}(t) = \frac{0.5 \cdot t^2 - 4.5 \cdot 10^{-6} \cdot t^3}{44 \cdot (2.5 \cdot 10^6 + 6 \cdot 10^{-3} \cdot t^2)}$

$$\mathcal{P}_{\text{apopt}}(t = 100) = 4.5 \cdot 10^{-5}$$

$$\mathcal{P}_{\text{apopt}}(t = 700) = 2.2 \cdot 10^{-3}$$

$$\mathcal{P}_{\text{apopt}}(t = 800) = 2.9 \cdot 10^{-3}$$

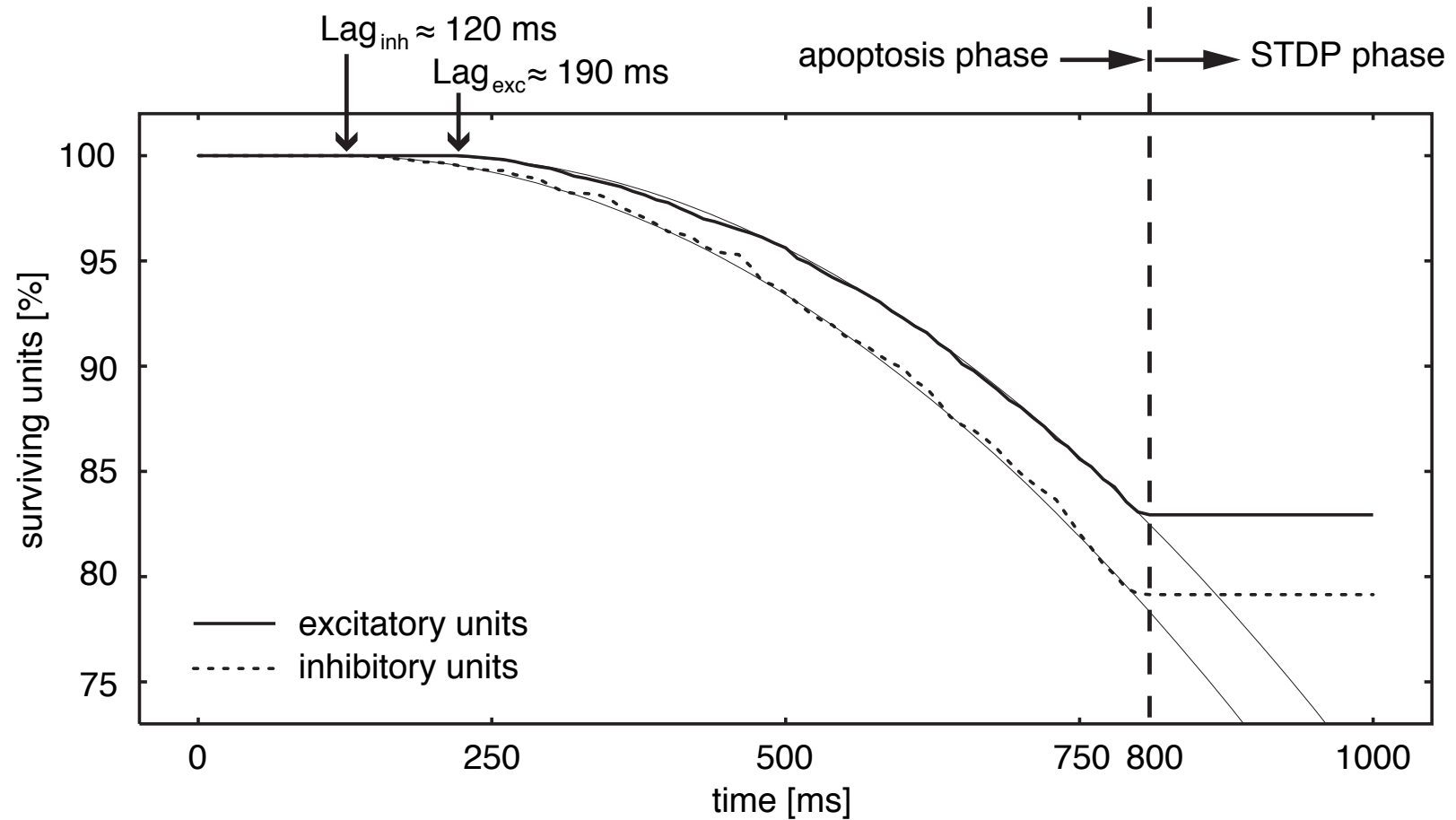
- **apoptosis induced by lack of excitatory afferents**

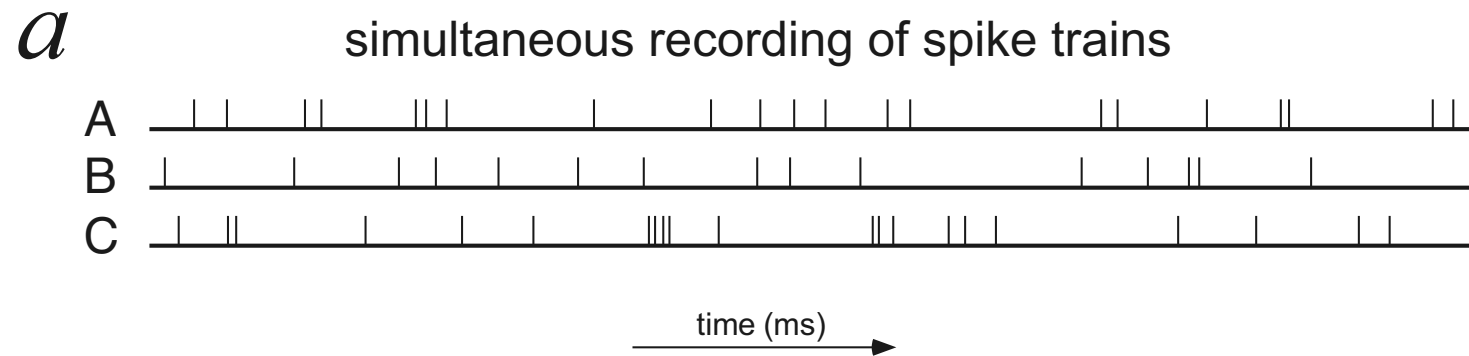
loss of all excitatory inputs due to:

apoptosis of pre-synaptic unit

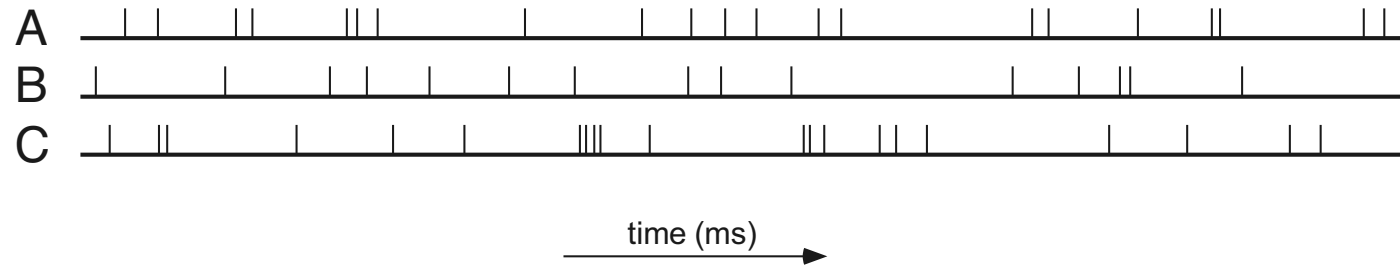
STDP driving to $A_{ji}(t) = 0$

- $0 \leq t < \{700, 800\}$ ms (initial phase)
apoptosis induced by excessive firing rate
- $\{700, 800\} \leq t < 10^5$ ms
Spike-Timing Dependent Plasticity
⇒ synaptic pruning
⇒ apoptosis induced by lack of excitatory inputs
- $t = \{1000, 3000, 5000, \dots\}$ ms
100 ms lasting stimuli
50 presentations (random mix: 50% AB and 50% BA)



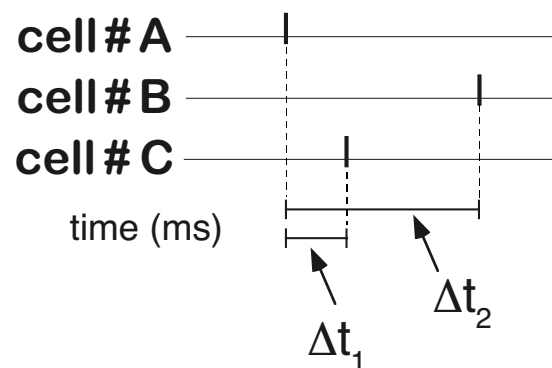


a simultaneous recording of spike trains



b detection of statistically significant spatiotemporal firing patterns

$\langle A, C, B; \Delta t_1, \Delta t_2 \rangle$



patterns found $n=3$

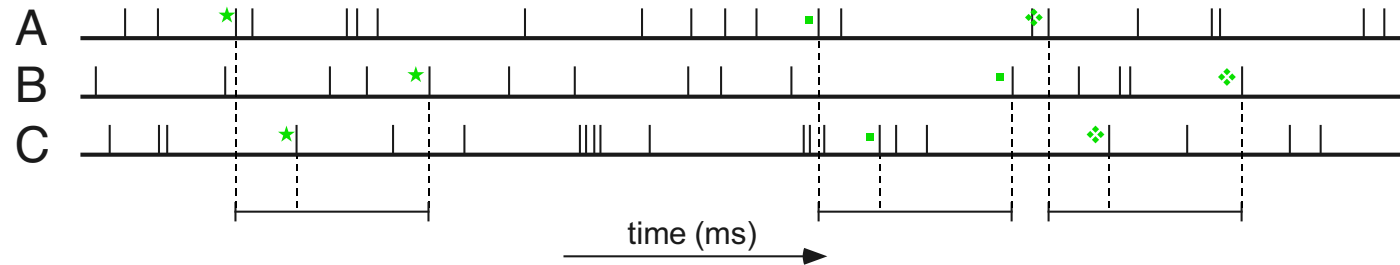
expected count $N=0.02$

significance of this pattern

$$pr(3, 0.02) \approx 1.3 \cdot 10^{-6} < 0.001$$

a

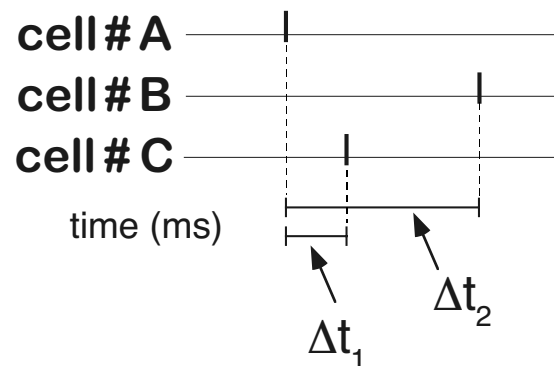
simultaneous recording of spike trains



b

detection of statistically significant spatiotemporal firing patterns

$\langle A, C, B; \Delta t_1, \Delta t_2 \rangle$



patterns found $n=3$

expected count $N=0.02$

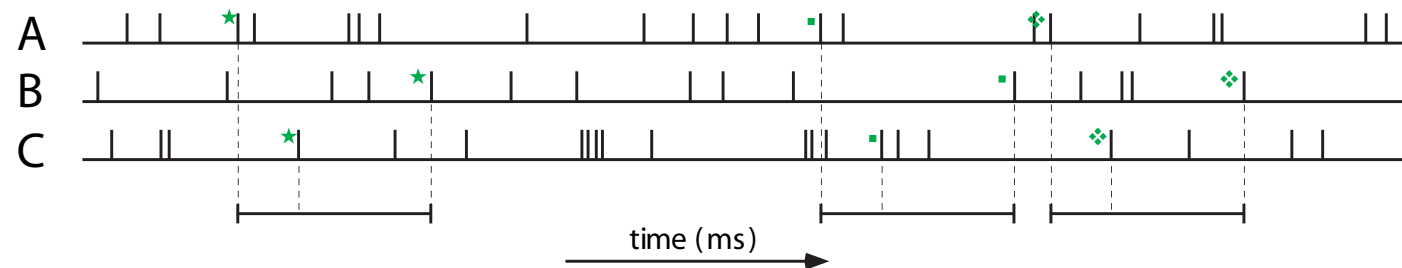
significance of this pattern

$pr(3, 0.02) \approx 1.3 \cdot 10^{-6} < 0.001$

for methods, see Villa *et al.*, 1999; Tetko and Villa, 2001

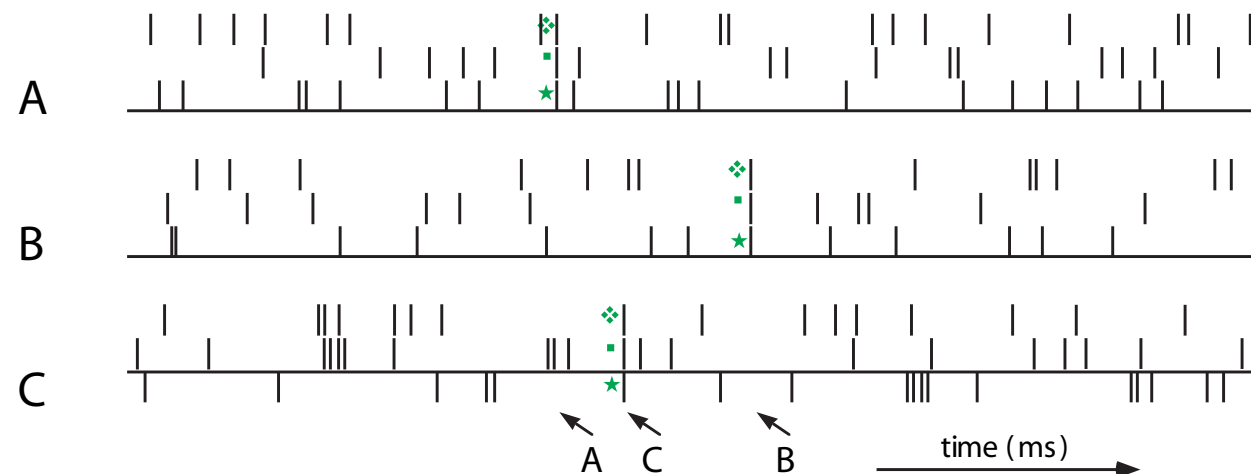
a

simultaneous recording of spike trains



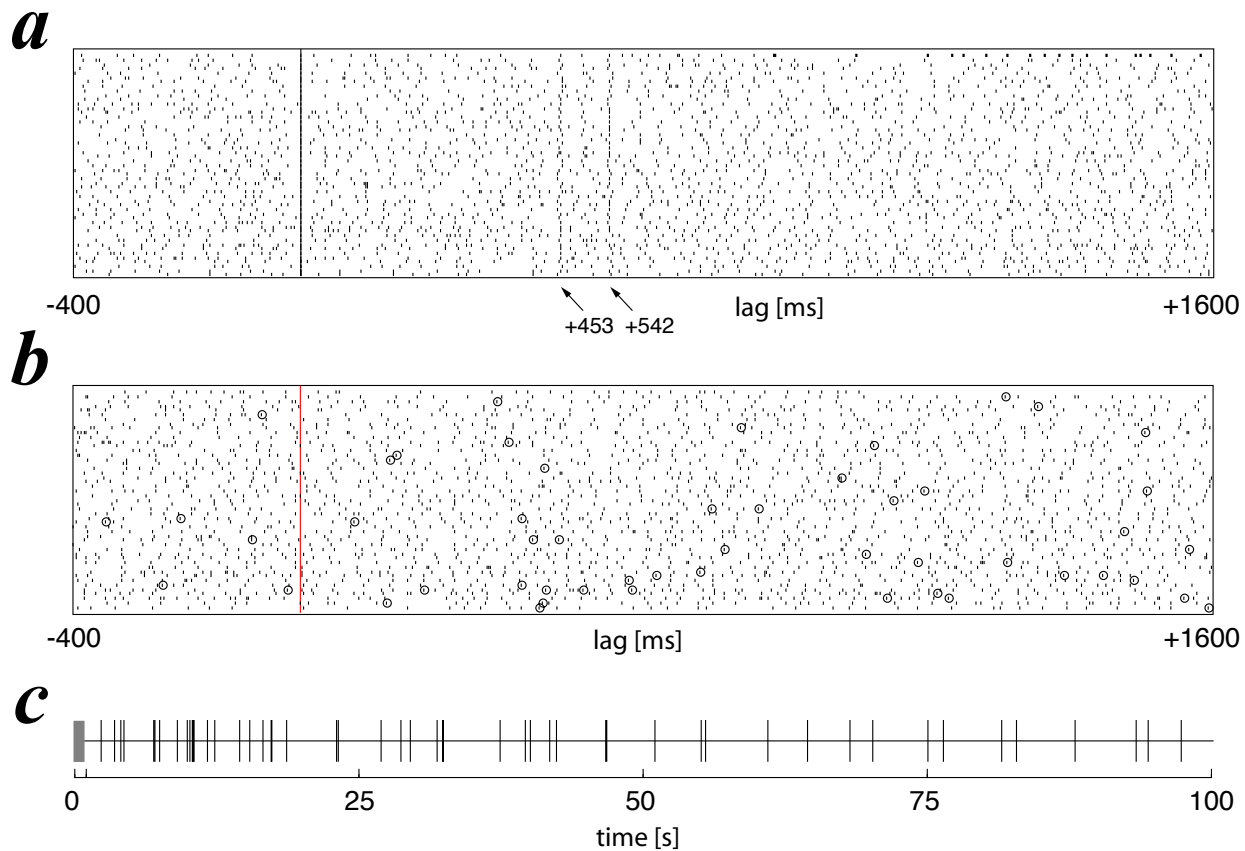
c

rasters of spikes aligned on pattern start



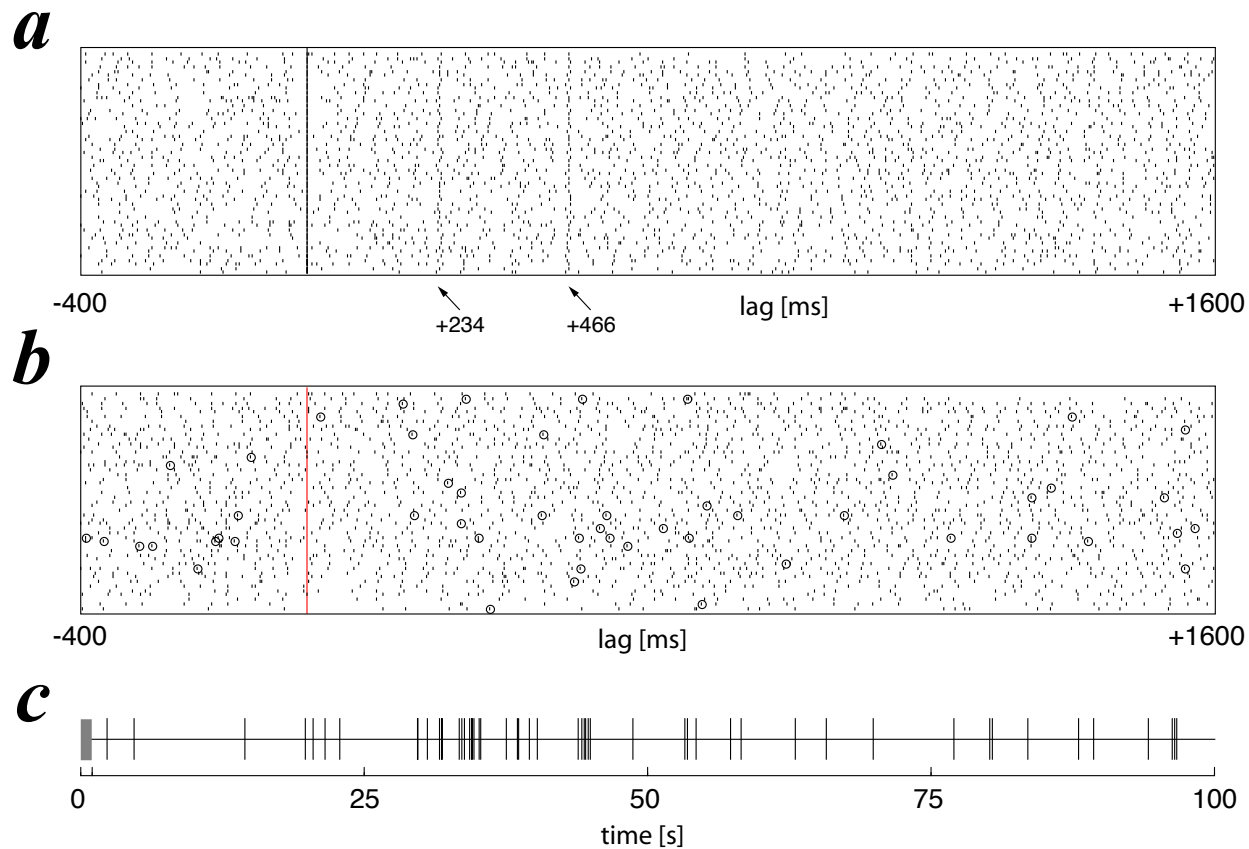
stopping firing rate-induced apoptosis at $t=700\text{ms}$

$\langle 79, 79, 79; 453 \pm 3.5, 542 \pm 2.5 \rangle$



stopping firing rate-induced apoptosis at $t=800\text{ms}$

$\langle 13, 13, 13; 234 \pm 3.5, 466 \pm 4.5 \rangle$



- inhibitory units enter apoptosis about 70 ms before excitatory units
- death dynamics of both populations followed the probability function to die $\mathcal{P}_{\text{apopt}}(t)$
- addition of cell death to the model improved stability of the network
 - stopping apoptosis at 700 ms lead to larger number of surviving units with lower mean firing rate
 - stopping apoptosis at 800 ms lead to smaller number of surviving units with greater mean firing rate
- maintained ability to let emerge cell assemblies associated to preferred firing sequences